

## ESTIMATING BACTERIAL PRODUCTION FROM $^{14}\text{C}$ -LEUCINE INCORPORATION INTO PROTEIN

In this lab, we will estimate bacterial production in your Winogradsky column by measuring the incorporation of "hot"  $^{14}\text{C}$  leucine into bacterial protein. To get a rate, we measure the amount of  $^{14}\text{C}$  incorporated at 0, 15 and 30 minutes. We halt the uptake by killing the bacteria with 100% trichloroacetic acid (TCA). TCA also precipitates proteins. We collect the bacteria by centrifugation and remove free unassociated  $^{14}\text{C}$  leucine by rinsing with 5% TCA and 80% ethanol. A scintillation cocktail will then be added that converts beta emission from  $^{14}\text{C}$  into light pulses by transferring energy given off by  $^{14}\text{C}$  disintegrations to fluor molecules that emit light at 320-420 nm. These bursts of light will be counted by a scintillation counter as CPM or DPM.

EVERYONE MUST WEAR A LAB COAT, COVERED SHOES AND GLOVES!

1. Label four 2 mL microcentrifuge tube *caps* "W", "0", "1", "2" which stand for "Working solution", Blank (0 min), 15 min incubation and 30 min incubation. Also place a small mark on the tube itself as well, which will allow us to place the tube in the centrifuge in the same orientation each time. Don't write on the side of the centrifuge tube (Why?)
2. Collect more than 5 mL of water from the top of your Winogradsky column and place in a snap cap tube. We actually only need 4.5 mL.
3. Add 20  $\mu\text{L}$  of the  $^{14}\text{C}$ -leucine working solution (which is 12  $\mu\text{M}$  Leu) to the bottom all four tubes. Keep all materials that come into contact with  $^{14}\text{C}$  on the tray at your lab bench.
4. Firmly cap and set the "W" sample aside, as we will not use it until the last step.
5. Add to the "0" tube ONLY, 0.1 mL of 100% cold TCA (trichloroacetic acid).
  - a. This is the killed control.
  - b. Leave on lab bench until incubation of other samples is completed.
6. To the "0", "1" and "2" tubes, add 1.5 mL of your Winogradsky water. Cap firmly, then shake sample. Be careful to NOT splash the  $^{14}\text{C}$ -leucine out of the tube.
  - a. This will give a final leucine concentration of 160 nM.
  - b. Do not touch the pipette tip to the liquid as it contains  $^{14}\text{C}$  and possibly TCA. If you do, replace the pipette tip.
  - c. NOTE THE TIME OF ADDITION!!
7. **The incubation period now begins.** In this case, just leave your samples on your tray in a rack. Incubations are typically done under conditions relevant to the samples.
8. After 15 min, kill sample "1" by adding 0.1 mL of 100 % cold TCA. Cap and shake.
9. After 30 min, kill sample "2" by adding 0.1 mL of 100 % cold TCA. Cap and shake.
10. Place all 3 samples on ice for 15 min; however, 30 min would be better but we do not have the time for this in today's lab.
11. Place your three samples in the centrifuge so that the mark on the tubes faces radially outward. NOTE the location of your samples in the rotor. **Make sure rotor is balanced with samples!!!**
12. Centrifuge samples on high for 15 min.

13. Remove tubes, and gently pore sample into  $^{14}\text{C}$  waste container. A cell "pellet" may, or may NOT, be visible on the outer wall of the centrifuge tube. Regardless of visibility, we do NOT want to dislodge this "pellet", so to not touch any pipette tip in the expected location.
14. Gently tap lip of tube on a folded Kimwipe placed on your workspace tray to removal all drops. Kimwipe needs to be treated as solid  $^{14}\text{C}$  waste. Also, avoid tapping tube on any drop collected on Kimwipe, as we do not want to contaminate the outside of the tube with  $^{14}\text{C}$ .
15. Add 1.0 mL of cold 5% TCA to each tube, but to not shake and avoid touching pipette tip on centrifuge tube or dispensing TCA onto phantom pellet.
16. Place tubes back in centrifuge, again with mark facing outward and noting location of your samples in the rotor.
17. Spin on high for 5 min.
18. Again, pour out liquid into sample waste, and gently tap tube on NEW Kimwipe to remove drops.
19. Add 1 mL of cold 80% ethanol to each tube.
20. Respin samples in centrifuge as before for 5 min.
21. Again, pour out liquid into sample waste, and gently tap tube on NEW Kimwipe to remove drops.
22. Place tubes in rack on your bench with caps OFF to allow samples to dry. You may need to come back after lunch for the final step.
23. Add 1.0 mL of scintillation cocktail to each tube, INCLUDING sample "W". HOWEVER, be very careful not to splash  $^{14}\text{C}$  leucine out from sample "W", and do not touch pipette tip to centrifuge walls.
24. Recap all samples and vortex for 10 sec or so.
25. Place all tubes in scintillation vials for counting. Place a mark on the top of each cent vial with your initials along with "W", "0", "1" or "2" as appropriate. Do not write on the side of the scintillation vials (why?).
26. We will take samples over for counting on the scintillation counter, which we will discuss at our next meeting.